Biomedical Applications of Cold Plasma

G DIVYA DEEPAK¹, ATUL²

(cc) BY-NC-ND

Review Article

ABSTRACT

Plasma medicine is branch that employs Cold Atmospheric Plasma (CAP) as a potent tool for biomedical applications. CAP produces high-level reactivity (free radicals, electrons) and can be generated by noble gases. CAP is rich in Reactive Nitrogen Species (RNS) and Reactive Oxygen Species (ROS). These ROS and RNS which include Ozone (O₃), Nitrous Oxide (NO), Hydroxyl Radicals (OH), and Nitrogen Dioxide (NO₂), gas particles, charged ions, neutral reactive oxygen which are primarily responsible for decontamination of microbes in various living tissues. Furthermore, CAP only have high excitation energies of electrons as compared to neutrals and ions which makes CAP an excellent tool for application on cells and tissues without any thermal damage. Cold plasma has also been successfully implemented for virus disinfection as its regarded as eco-friendly, efficient and novel technique for decontamination of virus. CAP treatment has also enabled inactivation of virus strain in both plant and animal species without inducing any physiological damage to them. Plasma chemistry essential for inactivation of pathogens is dependent on fine-tuning of various parameters which include plasma supply frequency, gas composition, input energy duration, pulse form, and modulation which has led to development and research of numerous portable plasma devices for different treatment methods in plasma medicine. CAP generated is extensively applied for wide range of biomedical applications including dentistry, microbial disinfection (bacteria, viruses, fungi), treatment of skin diseases, wound treatment, and biofilm treatment.

Keywords: Atmospheric pressure plasma, Disinfection, Plasma medicine, Reactive species, Sterilisation

INTRODUCTION

Plasma medicine comprises of Cold Atmospheric Plasma (CAP) to produce specific amounts of reactive species which are focussed on biological surfaces (tissues/cells). Plasmas have been used for a long time for sterilisation of medical equipment, packaging in the food industry, implants, blood coagulation, dentistry, biofilm treatment [1-6].

Cold atmospheric plasma has been implemented for wide spectrum of applications that include- implant surface treatment, medical equipment sterilisation, microbial disinfection. These applications are possible due to inherent benefits of CAP such as scalability, portability and disinfection effectiveness in small and confined spaces. Currently there are numerous CAP based devices that are employed for treatment of tissues and cells. CAP is implemented for rapid sterilisation. There are also new research possibilities for drugs delivery through tissues and biofilms [7-11]. The results of these specific experiments could translate into plasma based drug delivery techniques [12]. However, this would rely on factors such as conceptual design of plasma sources (whose chemical reactivity could be controlled), application of plasma physically, and most importantly in vivo and in vitro experiments.

The CAP is generated through several mechanisms that includemicrowave frequencies, Radiofrequency (RF), high voltage (DC/AC). Since, these CAP are non equilibrium plasmas, they consist of both reactive species (electrons and ions) as well as excited species that has immense potential in plasma based medicine as well as drug delivery [13-17].

Plasma medicine is an evolving field of research that is based fundamentally on plasma physics which governs the physical and chemical properties of CAP. The rapid development of CAP based techniques is attributed to the interdisciplinary nature of this research which encompasses various fields that include physics, chemistry, microbiology and engineering for characterisation, analysis as well application of this CAP technology. This review focusses on the current advancements and research pursued across these

domains. Numerous CAP based industrial applications are currently being employed in various engineering processes [18,19]. Even though, there are several existing applications of CAP in biomedical field and few efforts have been made to understand the impact of CAP on biofilms (complex microorganism groups enclosed by glue matrix which further improves resistance towards external stresses) [20]. As the conventional antimicrobial therapy has poor penetration capacity it is not particularly effective against the biofilms which is primarily responsible for inhibiting disease eradication. Also, the conventional antimicrobial therapy several drawbacks like systemic toxicity and bacterial resistance. But when the bactericidal effects of CAP are considered, then it is found that the resistance of bacteria remains the same while being exposed to CAP. These results are clearly suggested that CAP is potent tool for in-vivo treatments [21]. Even though, CAP is generally close to room temperature, yet it contains sufficient chemically reactive gas species utilised for

contains sufficient chemically reactive gas species utilised for numerous biomedical applications. In this CAP only electron possess high excitation energies as compared to the ions and neutrals. Furthermore, during the CAP generation process these high energy electrons collide with the working gas (He/Ar/N₂/O₂) which is used for generating plasma thus resulting in higher ionisation and disassociation levels. As the neutrals and ions are comparatively at a much lesser temperature than electrons in the CAP hence there is no thermal damage. It is imperative to note that this property enables CAP to be implemented for tissues, cells, biological matter, thermally sensitive materials, [22-24].

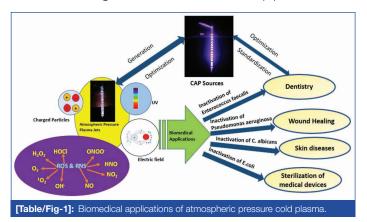
Recently, huge number of CAP devices have been envisaged, designed, developed and implemented for many research related applications. Broadly, these non equilibrium CAP have been classified as indirect and direct discharge [13-17]. Since, various research groups use different CAP based devices for numerous biomedical applications thus making the comparison between them quite complicated. Due to these technical differences of design of various CAP devices developed across globally it becomes imperative to implement a standardised way to comprehend the effectiveness and

security of the CAP devices. For this specific reason, there are many organisations {European Committee for Standardisation (CEN), German Institute of Standardisation (DIN), International Organisation for Standardisation (ISO)} for technical regulation of the CAP device [25]. Some of the CAP devices have already got the approval of United States Food and Drug Administration to for the clinical usage.

The CAP applications have received lot of attention from the medical research community. The biomedical applications of CAP include, viral infection treatment (herpes simplex), wound treatment and skin diseases, implant surface treatment, biofilm treatment etc.

PLASMA MEDICINE

The CAP have been employed in wide spectrum of biomedical applications that are depicted in [Table/Fig-1] (conceptualised by the authors) which includes dental medicine, treatment of skin diseases and wound healing and sterilisation of medical equipment.



It is essential to perform modelling, simulation and experiment work on these atmospheric pressure plasmas for understanding the chemical and physical mechanisms in detailed manner. Certain important parameters such as seed electrons, photo ionisation effect and impact of electric field can be obtained using fast imaging techniques that have high resolution up to micrometre range and time resolution of up to nanoseconds.

For examining the active components of CAP which include Reactive Oxygen Species (ROS), Reactive Nitrogen Species (RNS), electric fields, and Ultraviolet (UV) radiation, their generation mechanism as well as their interrelationship between them should be examined in detailed manner.

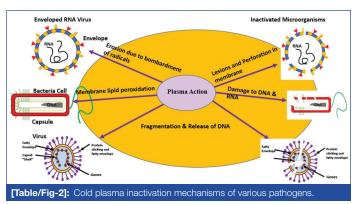
Numerous existing measurement technology are employed for studying CAP which include- optical emission spectroscopy for gas discharges, Rayleigh and Raman for understanding electron parameters, mass spectroscopy, stark spectroscopy. The plasma reactive species generation mechanism could be understood through modelling and simulation of various plasma reactions [26].

When CAP based devices have also been used for bioaerosols and fluids also, thereby plasma interacts with water directly. The lifespan of the bioaerosols relies on the evaporation of water and thereby impacts the plasma chemistry. While implementing CAP for treatment of skin diseases, it essential to adjust the treatment plasma dose accordingly [27]. CAP sources scale from small to large size devices and their design depends on several factors such as surface type of target, electrical parameters for a particular biomedical application (voltage, current, supply frequency, pulse width). The rapid progress in the field of CAP technology is moving towards incorporating it in personalised devices, laptops and other portable devices including those operating on battery.

The implementation of Artificial Intelligence (AI) in plasma diagnostics and mathematical modelling of plasma reactions would lead to rapid progression in commercialisation of CAP based devices with augmented disinfection efficacy [28]. Furthermore, innovative highstrength materials can used as electrode materials, semiconductor based power sources can be used to increase efficiency of CAP devices while also maintaining cheap manufacturing cost.

COLD PLASMA APPLICATIONS- INACTIVATION OF PATHOGENS

Some microorganisms such as viruses, bacteria, and fungi behave as pathogens and trigger diseases. The resistance of these microbes is quite different from each other and can be classified as very susceptible, less resistant, intermediate resistant, highly resistant, and most resistant [29]. The various cold plasma inactivation mechanisms for pathogens are shown in [Table/Fig-2] (conceptualised by the authors).



Helminth eggs, protozoan oocysts, and bacterial spores fall in category of microorganisms which are highly resistant. The microbes such as fungal spores, protozoan cyst and non enveloped viruses fall in category of intermediate resistant microbes. The type of cold plasma treatment is different for various types of microbes and depends on the resistance of the microbe [30].

Bacterial Inactivation-Cold Plasma

Numerous studies that have explored the advantages of cold plasma in treating highly drug resistant bacteria [30-35]. Various studies have also proved that cold plasma produces Reactive Oxygen and Nitrogen Species (RONS) when using air as working gas [36,37]. The investigation done by Nicol MJ et al., proved that there was about 90% reduction in both gram negative and gram positive strains of bacteria on different surfaces (solid, porous, liquid) [38]. Furthermore, these experiments demonstrated the generation of RONS during cold plasma treatment is essential for inactivation of bacteria.

The diverse benefits of CAP which include portability, scalability and efficacy of the technology has resulted in numerous microbial disinfection-based applications. As this cold plasma is generated at room temperature hence can be applied to biomaterials without any thermal damage. Another benefit of CAP treatment it's not dependent on liquid chemicals and relies on dry chemistry. Furthermore, results of numerous investigations have proved that bacterial growth has not taken place after CAP treatment [39-41]. CAP treatment has been extensively applied for inactivating bacteria in biofilms is dependent on bacterial cell wall thickness. Few studies also suggested the efficacy variation of CAP treatment for gram negative and gram positive bacteria due to their cell thickness difference [40-42]. The experimental results of Mai-Prochnow A et al., showed that gram negative bacteria (Pseudomonas aeruginosa) having cell thickness of 2.4 nm was completely inactivated using CAP treatment whereas in case of gram positive bacteria (Bacillus subtilis) with a 55.4 nm cell wall thickness had the highest resistance to CAP treatment [42]. The inactivation of bacteria is known occur via three different mechanisms which include: (i) Deoxyribonucleic Acid (DNA) direct chemical damage; (ii) Permeabilisation of cell membrane causing cell component leakage; (iii) Damage to intercellular proteins via nitrosative or oxidative species [43]. Cells go through a sequential set of morphological and physiological changes before being inactivated.

The experimental results of Kvam E et al., showed that the CAP treatment had damaged the outer membrane and cell wall of gram negative bacteria [44]. Recently, the cold plasma has acquired considered attention due to its efficacy in inactivation of surface contaminants. Joshi SG et al., had successfully designed Floating-Electrode Dielectric-Barrier Discharge (FE-DBD) for swift inactivation of bacterial contaminants in air [45]. Joshi SG et al., validated that the *E. coli* membrane lipid peroxidation can be done using FE-DBD [46].

The response of bacteria towards the CAP treatment depends on species type. It is observed that gram negative bacteria comparatively more sensitive than gram positive bacteria, which indicates that the impact of CAP treatment depends on cell wall thickness as well as cell membrane. The experimental results of Laroussi M et al., proves that CAP treatment had induced considerable damage to gram negative bacteria. Though, the outer membrane is absent in gram positive bacteria (*B. subtilis*) it is highly resistant due to its thick cell wall that ensures its rigidity. Further, even though cell structure of *B. subtilis* was not affected due to CAP treatment but cell viability reduced drastically [47].

The CAP treatment is used widely in disinfection of microbial biofilms. Biofilm is regarded as cell clusters of bacteria encapsulated via Extracellular Matrix (ECM) [48]. Biofilm cells have been shown to display greater drug resistance as compared to planktonic cells [49]. The existence of ECM in biofilms significantly impacts the efficacy of CAP treatment. The matrix consists of 3-dimensional biofilm structures wherein cell to cell communication is done via embedded cells. The matrix is different for various bacterial species. The matrix comprises of nucleic acids, polysaccharides, proteins and Extracellular Polymeric Substance (EPS) [50].

The EPS serves as good barrier for chemicals which include biocides, antibiotics and provide a shielding effect for photons, RONS, charged particles generated in CAP during treatment. Hence, longer treatment duration is essential for mediation of EPS structure and thereby imitating enzymatic degradation effect. Once the reliability of biofilm matrix is damaged then the biofilm cell in the interior is more vulnerable to inactivation [51]. The ECM is not the only factor that is accountable for variable killing time of biofilm cells but there are other factors like oxidative stress response to planktonic cells thereby enabling them to be highly resistant to antibiotics [52].

Previous studies also indicated the RONS is concentrated in microcolonies [53,54]. But low doses of oxidative stress found to have favoured the growth of biofilms (*S. aureus, E. coli*). *S. aureus* stimulates biofilm formation in an oxidant dependent manner also biofilm formation improves when it is exposed to hydrogen peroxide. Hence, the CAP treatment duration and dose must be carefully chosen to make sure eradication of only the biofilm cells without damaging the other cells.

Biofilm growth occurs mostly often in moist environment, hence it is imperative to decode the ambient air as well as interactions of CAP with liquid based surfaces. Numerous studies have proved that efficacy of CAP treatment is significantly higher in wet surfaces as compared to dry surfaces. Reactive species generated during CAP treatment penetrate via diffusion through liquid based medium [55,56].

In spite of various benefits of CAP treatments in liquid based medium, there are a few drawbacks such as production of numerous chemical species (nitrate, hydrogen) and generates acids [57]. The antibacterial effect of CAP on liquid based media depends on the reactive species generated during treatment [58]. Radicals also play a main role in fatty acid oxidation inside the bacterial cell thereby inactivating the microbe [59]. The inactivation time of microbes relies on the liquid volume [60]. Thus, it is seen that bactericidal effect of CAP treatment in liquid interfaces is due to the generation of RONS.

Virus Inactivation-Cold Plasma

Virus is one of the major microbes that have infected various cellbased living organisms that include- humans, plant, animals. Most of these virus strains have significantly impacted the medical, agricultural sectors both biologically as well as economically. Cold plasma implemented for virus disinfection is regarded as ecofriendly, efficient and novel technique for decontamination of virus. Most of these virus strains are not dangerous and few of them are much beneficial for their host organism [61,62]. Various chemical (alkalis, chlorine, alcohols, and bleach) and physical methods (filtration, temperature, UV irradiation, pressure) have been employed for killing viruses [62]. The disinfection method differs based the matrix type and virus type considered for inactivation. Plant virus (tobamoviruses) and waterborne viruses (enteric viruses) are most resistant of all viruses [63,64]. Strong disinfection methods are essential for disinfecting stable viruses and it imperative to ensure it maintains the quality of water with increasing in toxicity. Traditionally, chlorination has been implemented for decontamination of water and it's observed that it's inefficient in killing some virus strains along with increasing the toxicity of water resulting in a health hazard.

Recently, numerous inactivation techniques have been developed which includes-UV, ozone treatment, membrane filtration [64]. The intriguing composition of CAP which includes reactive species, molecules, radicals and charged species has resulted broad research on the implementation of CAP for decontamination of pathogens.

Enteric viruses: CAP treatments have mostly concentrated on enteric viruses such as adenovirus, hepatitis A virus and norovirus. These viruses have been the prominent cause of infectious diseases globally [65]. Experimental research work with human viruses poses significant health risk and such investigations need latest technology labs and equipment. Due to limited cultivation methodologies the impact of enteric viruses (norovirus) has been analysed only with limited research data. Owing to these limitations they are mostly replaced via surrogate viruses.

Animal viruses which include Tulane virus, Feline Calicivirus (FCV) and murine norovirus are implemented as surrogates for norovirus due to their similarity about genetics, similar sizes and morphologies. Due to their augmented safety and easier to reproduce these surrogate viruses are being implemented. Furthermore, few of viruses have also been found to infect bacteria known as bacteriophages, these can also be implemented as surrogates [66].

The surrogates and enteric viruses have been successfully treated in various mediums that include stainless steel, food surfaces, liquids, aqueous solutions [67-70]. It was distinctly observed that the efficacy of CAP treatment was significant in surrogate virus as compared to the enteric virus [69,70] which clearly suggests the experimental results of CAP treatment with surrogates cannot be directly into consideration for understanding the enteric virus [69,70].

The FCV inactivation in liquid (including bacteriophages) has been implemented by CAP treatment with duration of 15 s [71,72]. Though, its short period of treatment time this suggests that CAP treatment is most essential for inactivation of virus in liquid based medium.

The CAP treatment has been implemented on various types of food surfaces which include- meat, lettuce, blueberries wherein the viruses have been successfully inactivated without modifying the physical or chemical properties of food. Furthermore, dielectric barrier discharge based CAP treatment has been applied packaged food also [68,69,73,74]. As these are direct plasma treatment methods, the temperature at the point of treatment should kept within permissible limit to avoid thermal damage of food in certain conditions due to various generation CAP generation techniques. Indirect CAP treatment methods like plasma-activated liquids can also be used for microbial decontamination. **Respiratory viruses:** Respiratory infections triggered by viruses are most important sources of upper and lower respiratory tract diseases. Among these the paramyxovirus and influenza virus are the major microbes. These viruses (influenza virus and paramyxovirus) are predominantly transmitted through air and by droplets of infected people.

One of the vital contents of CAP is ozone which is responsible for inactivation of viruses. The experimental results of Murray BK et al., proved that ozone triggers the peroxidation of proteins and lipids which in turn kills the virus [75]. Pulsed power based CAP treatment has been already implemented on Respiratory Syncytial Virus (RSV) and influenza virus [76]. In paediatric medicine this RSV is considered one of the most important virus as its transmission occurs via contaminated surfaces [77]. CAP treatment has also been implemented for efficient disinfection of various hospital devices (scalpels, dental instruments, clamps) [78]. As most of respiratory infections are airborne and since viruses causing them are very stable so it's imperative to sterilise the air to stop the virus transmissions which are airborne. The investigations done by Wu Y et al., and Xia T et al., showed successful eradication of MS2 bacteriophage using CAP treatment in duration of only 0.12 s and 0.25 s of contact time with CAP [79,80]. However, these CAP treatment techniques can at times be hazardous as ozone is produced in high concentrations. Thus, the CAP treatment employed for biomedical applications should be having ozone concentrations within the recommended safety limit [81].

Animal viruses: The most important viruses inactivation through CAP treatment are: Newcastle Virus (NV), Avian Influenza Virus (AIV), Porcine Reproductive and Respiratory Syndrome (PRRSv). These viruses impacted the food security and the economy. Some strains of NV can result in up to 100% mortality in bird species. Vaccines serve as the potent tool in offering better immunity via virus inactivation thereby reducing potential risk of disease [81]. Since, virus inactivation is preliminary step in vaccine preparation hence CAP treatment becomes quintessential to achieve the same.

Furthermore, there are few methods in which aqueous solutions activated by CAP treatment are employed successfully to disinfect tools and surfaces infected with poultry. Su X et al., proved that CAP activated aqueous solutions had completely inactivated viruses of avian embryos which attained 100% survival rate [82]. PRRSv is one the major microbes that has significantly impacted the pork industry. PRRSv is airborne virus which remains active even after traversing long distances [83].

Few of the most extensively employed methods for purifying air depends on either reducing virus contamination (different filters) or virus spread (UV irradiation). CAP treatment offers numerous advantages like curbing the spread of virus and killing them via charge driven filtration and RONS in CAP [84].

Plant viruses: Plant viruses are known to be predominantly transmitted via insects. These plant viruses are known to cause huge damage to crop globally resulting in economic downfall. The viral spread has been rapid due to various factors like closed irrigation system and untreated waste water [85]. In spite of this there are only few reports of CAP treatments inactivating these viral pathogens. which include water transmissible virus and potato virus [86,87]. Some of these economically significant plant viruses (Tobacco Mosaic Virus) (TMV) are found to be highly resistive and stable to various traditional inactivation methods. The experimental results of some studied showed DBD based CAP treatment for 10 minutes reduced the viral particles to subunit level and damaged the TMV Ribonucleic Acid (RNA) and thus successfully stopped spread of infection. Although, only few studies have been conducted for analysing the effect. Min SC et al., had successfully proved the inactivation of Tulane virus in roman lettuce using a DBD based CAP treatment [69]. These experimental results strongly suggests that plant viruses can be inactivated using CAP treatment without inducing any physiological damage to the plant species.

Fungi Inactivation-Cold Plasma

The CAP treatment is potent tool implemented in augmenting food safety and enhances shelf life. Fungal food spoilage has been a notable concern in the agriculture sector. Many of fungal infections once commenced is very tricky to disinfect as spores and fungal cells are known to demonstrate quite resistant structures. For inhibiting fungal spoilage of numerous fruits specifically citrus fruits. The traditional disinfection methods include wax coating, chemicals fungicides, [88].

To combat food security and satisfy the ever-growing consumer demands. Various CAP treatment technologies have been the scope of research in the agriculture sector.

Fungi are microbes which single celled or complex multicellular micro-organisms and nucleated. Fungi is further subdivided into different categories that include: (i) macroscopic filamentous fungi; (ii) multicellular filamentous moulds; (iii) single cell yeasts. Moulds comprise of fine threads known as hyphae. Hyphae branch out and form mycelium (network of threads).

Hayashi N et al., demonstrated the efficacy of atomic oxygen in killing *Penicillum digitatum* and *Aspergillus oryzae* [89]. Suhem K et al., had successfully implemented a RF-based plasma which inactivated *Aspergillus flavus* [90].

Avramidis G et al., applied atmospheric pressure DBD plasma treatment for disinfecting *Ascochyta pinodella* and *Fusarium culmorum* fungi [91]. After the CAP treatment it was observed that the cell membranes and cell wall structures were significantly affected which led to cytoplasm leakage [91].

Lee GJ et al., demonstrated that CAP treatment on Cordyceps bassaiana had significantly damaged its fungal spores causing wrinkling of surface, shrinkage, flattening and rupture and modified the fungal spores morphologically [92]. Experimental results of Dasan BG et al., showed that fungal spores of *Aspergillus parasiticus* were eradicated using CAP treatment and it also revealed the cell constituents scattered in clusters [93].

Extensive investigation on the impact of 1O2 (singlet oxygen) in killing fungi has been done by Hashizume H et al., [93-96]. Iseki S et al., employed atmospheric pressure plasma consisting of high density ground state atomic oxygen for inactivation of *Penicillium digitatum* fungal spores [97]. Eisenman HC and Casadevall A further proved that fungal spores consist of melanin (protective pigment) in the cell wall structure that shields it form external parameters such as UV radiation [98].

Thus, it can be concluded that CAP treatment causes leak of intracellular composition by damaging the cell wall structure of the fungi spores. It is imperative to note that magnitude of changes in fungi spores is directly proportional to its exposure to reactive plasma constituents (OH, H_2O_2 , HNO₂, H, O, e-).

COLD ATMOSPHERIC PLASMA TREATMENT

CAP Treatment for Fungal Disinfection of Food

The CAP treatment has been extensively implemented for a wide variety of food stuffs that include- food grains, dried meat, nuts, fish, spices and herbs [99-106]. Whitehead indicated that plasma chemistry is specifically dependent on fine-tuning of various parameters which include plasma supply frequency, gas composition, input energy duration, pulse form, and modulation [107]. Also, CAP treatment is much preferred over other traditional decontamination techniques as it does not employ intense system [108-110]. Furthermore, CAP treatment eliminates pathogens in food stuff without causing any thermal damage to food products [111].

CAP Treatment in Wound Healing

The CAP treatment has played a pioneering role in wound healing. Preliminary investigations and experimental studies prove that CAP are very efficient in enhancing healing of chronic wounds in both humans and animals without causing any considerable damage to healthy tissues [112-116]. These experimental results also showed that CAP treatments reduced the bacterial load substantially [117]. The investigations of Lou BS et al., established that CAP has the capability to disinfect microbes by damaging their DNA and via cell wall destruction thereby reducing the microbial load [117]. In addition to microbial disinfection the CAP treatments were also found to be instrumental in alteration of inflammation of chronic wound which transformed prevented the stagnation of wound [117]. Wound healing predominantly concerned with cell migration and proliferation as well as angiogenesis. Mainly, fibroblasts and keratinocytes are the cell types which enables wound healing, amongst which keratinocytes is played a key role in the main healing processes whereas the fibroblast cells played a guiding role [118]. Furthermore, the CAP treatments improved numerous growth aspects (neovascularisation/angiogenesis) along with reactive plasma species interaction (atomic oxygen, OH, NO) [118-121]. The CAP generates RONS that are capable for healing wounds and may accelerate the signalling paths to normalise tissue healing in the skin [122]. It is important to note that certain biological mechanisms such as cell proliferation and migration and tissue repair have been found to be accelerated by impact of RONS [123]. Ngo MHT et al., found that fibroblast growth was responsible for RONS induced endothelial cell proliferation [124].

The CAP due its distinctive characteristics and constituents have been widely employed in numerous biomedical applications such as wound healing, normalise drug resistant bacteria, and root canal treatment [125,126]. However, a numerous reports claimed that UV radiation in plasma is low and does not play a significant role in anti microorganism process except that from microwave driven discharge [127-130]. Furthermore, it is important to note that the plasma dose, gap between plasma source and specimen, duration of treatment is essential to optimise the efficacy of CAP treatment [131].

CAP Treatment in Dentistry

One of the most important advantages of CAP treatment is its ability to sterilise and treat irregular surfaces. CAP treatment in dentistry involves the reactive species in plasma interacting with internal dental cavity. Sladek REJ et al., demonstrated that plasma needles effectively inactivated Escherichia coli [132]. The results of Goree J et al., proved that CAP treatment effectively disinfected Streptococcus mutans [133]. Sladek REJ et al., examined the plasma interactions with dental tissue implemented via plasma needle [132]. Since, CAP treatment operates at room temperature it does not cause bulk destruction of the tissue. Furthermore, the sterilisation and cleaning of infected tissue in dental cavity can be implemented via laser or mechanical technique. In both these techniques there can be damage of healthy tissue as well as heating issue. To overcome these challenges the plasma needle is employed as efficient source containing reactive species to accomplish the microbial disinfection without harming the healthy tissue. The reactive species in gas phase produced by the plasma needle can interact on tooth surface and can dissolve into a liquid. Plasma needle generates bactericidal agents in treatment vicinity and penetrates fissure spaces and internal part of the dental cavity [134]. Yang B et al., designed plasma brush using argon as working gas. This plasma brush was found be highly efficient in decontamination and inactivating microbes such as Streptococcus mutans and Lactobacillus acidophilus [135]. Their experimental results suggested 100% bacterial disinfection within treatment time of 15 seconds for Streptococcus mutans and within five minutes for Lactobacillus acidophilus.

Thus, it can be summarised that CAP treatment is novel technology capable of microbial disinfection which can be implemented in narrow channels as well as irregular structures. Adhesive restoration: Dong X et al., examined the effects of CAP treated dentin surfaces and their interaction with 2-Hydroxyethyl Methacrylate (HEMA), adhesive monomer [136]. Their experimental results clearly showed that argon based CAP treatment was very effective in enhancing adhesive monomer penetration and subsequently enhanced the dentin/adhesive interface bonding. Ritts AC et al., implemented CAP based brush for composite restoration [137]. Their results demonstrated that CAP treatment enhanced the surface characteristics of dentin which subsequently improved the bonding between adhesive restoration and dentin. Furthermore, CAP treated fiber-reinforced composite and resin were found to demonstrate better tensile shear bond as compared to traditional core [138].

Biofilms: Biofilms formed on the tooth surface results in periodontal diseases, dental cavity, oral mucositis and periodontal diseases. These biofilms also impact the dental implant resulting in conditions such as peri-implantitis and peri-mucositis. Furthermore, CAP treatment has the capacity penetrate and destroy biofilm matrix without damage to the oral tissue [139]. The study done by Koban I et al., found the CAP treatment has more efficacy in destroying the microbes (bacteria) present in dental biofilm in comparison to chemical antibacterial agents such as chlorhexidine [140]. Jiang C et al., implemented a plasma plume to decontaminate the root canal effectively at room temperature [141].

Tooth whitening: The CAP treatment has been widely implemented in tooth bleaching. Lee HW et al., demonstrated helium based CAP jet in combination with hydrogen peroxide solution (as catalyst) can be effectively used for tooth bleaching [142]. Their experimental results suggested that tooth bleaching effect generated by helium based CAP jet was due enhanced OH production and removal of tooth surface proteins. Nam SH et al. studied the effectiveness of tooth bleaching using CAP treatment (with 15% carbamide peroxide (CP; CH_aN₂O₂) including 5.4% Hydrogen Peroxide (HP), in comparison to other traditional light sources. Their results proved that CAP treatment was better and efficient in tooth bleaching without any resulting in any thermal damage as compared to conventional light sources (low concentration of hydrogen peroxide). Furthermore, direct current plasma jets (with hydrogen peroxide) have also been successfully implemented in tooth whitening [143]. The removal of intrinsic stains has presented big challenge during teeth bleaching [144]. Park JK et al., implemented a low frequency plasma source (used with hydrogen peroxide) for eliminating intrinsic stain [145]. The in-vitro investigations of Kim MS et al., showed that CAP treatment could be used for tooth bleaching [146]. They also found that the plasma treatment had not resulted in any damage to the tooth.

The CAP treatment has created an intriguing new era of dental care. The CAP treatment efficacy in various domains of dentistry (root canal disinfection, sterilisation, tooth whitening) has been encouraging for its wide implementation. As the oral disorders are polymicrobial nature, hence it's imperative to understand inactivation mechanism of CAP for each of these microbes responsible for dental plaque development. Further, there is research lacuna which needs to be investigated for understanding the CAP interaction with living tissues/cells.

CONCLUSION(S)

The CAP has been implemented as potent tool in wide spectrum of biomedical applications (food sterilisation, wound treatment, dentistry). It is seen that CAP is also very effective in inactivation and decontamination of microbes (bacteria, viruses, fungi). CAP treatment has also successfully inactivated food borne microbes responsible for food wastage. Furthermore, CAP has also been increasingly implemented for food processing techniques which enhance food quality (functional, sensory, texture properties). As CAP treatment is done at room temperature the thermal damage to the food or tissue is avoided. It also be noted CAP based medical devices have also been tested and implemented for various biomedical applications. Thus, CAP has shown tremendous potential for its effectiveness in biomedical without modifying/damaging surface properties of tissues/cells.

REFERENCES

- [1] Deng S, Ruan R, Mok C K, Huang G, Lin X, Chen P. Inactivation of *Escherichia coli* on almonds using nonthermal plasma. J Food Sci. 2007;72:M62-66.
- [2] Deilmann M, Halfmann H, Bibinov N, Wunderlich J, Awakowicz P. Low-pressure microwave plasma sterilisation of polyethylene terephthalate bottles. J Food Prot. 2008;71:2119-23.
- [3] Fridman G, Friedman G, Gutsol A, Shekhter AB, Vasilets VN, Fridman A. Applied plasma medicine plasma process. Polym. 2008;5:503-33.
- [4] Moreau M, Orange N, Feuilloley MGJ. Non thermal plasma technologies: New tools for biodecontamination. Biotechnol Adv. 2008;26:610-17.
- [5] Selcuk M, Oksuz L and Basaran P. Decontamination of grains and legumes infected with Aspergillus spp. and Penicillum spp. by cold plasma treatment Bioresour. Technol. 2008;99:5104-09.
- [6] Deng X, Shi JJ, Kong MG. Protein destruction by a helium atmospheric pressure glow discharge: capability and mechanisms J Appl Phys. 2007;101:074701.
- [7] Lerouge S, Wertheimer MR, Yahia L'H. Plasma sterilisation: A review of parameters, mechanisms, and limitations. Plasmas Polym. 2001;6:175-88.
- [8] Moisan M, Barbeau J, Moreau S, Pelletier J, Tabrizian M, Yahia L'H. Lowtemperature sterilisation using gas plasmas: A review of the experiments and an analysis of the inactivation mechanisms. Int J Pharm. 2001;226:01-21.
- Laroussi M. Non thermal decontamination of biological media by atmospheric pressure plasmas: Review, analysis and prospects IEEE Trans. Plasma Sci. 2002;30:1409-15.
- [10] Sharma A, Pruden A, Zengqi Y, Collins GJ. Bacterial inactivation in open air by the afterglow plume emitted from a grounded hollow slot electrode Environ. Sci Technol. 2005;39:339-44.
- [11] Sladek RE, Stoffels E. Deactivation of Escherichia coli by the plasma needle. J Phys D Appl Phys. 2005;38:1716-21.
- [12] Laroussi M, Mendis DA, Rosenberg M. Plasma interactions with microbes. New J Phys. 2003;5:41.
- [13] Divya Deepak G, Joshi NK, Pal UN, Prakash R. Electrical characterisation of atmospheric dielectric barrier discharge-based plasma cold plasma jet using ring electrode configuration. Laser and Particle Beams. 2016;34:615-20.
- [14] Divya Deepak G, Joshi NK, Pal DK, Prakash R. A low power miniaturized dielectric barrier discharge based atmospheric pressure plasma jet. Review of Scientific Instruments. 2017;88:013505.
- [15] Divya Deepak G, Joshi NK, Prakash R. Modal analysis and electrical characterisation atmospheric pressure cold plasma jet in pin electrode configuration. AIP Advances. 2018;8:055321.
- [16] Divya Deepak G, Joshi NK, Prakash R. Electrical characterisation of argon and nitrogen based cold plasma jet. Eur Phys J Appl Phys. 2018;83:20801.
- [17] Divya Deepak G, Joshi NK, Prakash R. Modal analysis of dielectric barrier dischargebased argon cold plasma jet. Laser and Particle Beams. 2020;38:229-38.
- [18] Brandenburg R, Ehlbeck J, Stieber M, Woedtke TV, Zeymer J, Schlüter O, et al. Antimicrobial treatment of heat sensitive materials by means of atmospheric pressure Rf-driven plasma jet. CoPP. 2007;47:72-79.
- [19] Brandenburg R, Lange H, Woedtke TV, Stieber M, Kindel E, Ehlbeck J, et al. Antimicrobial effects of UV and VUV radiation of nonthermal plasma jets. IEEE Trans Plasma Sci. 2009;37:877-83.
- [20] Laroussi M. Low-temperature plasmas for medicine? IEEE Trans Plasma Sci. 2009;37:714-25.
- [21] Delben JA, Zago CE, Tyhovych N, Duarte S, Vergani CE. Effect of atmospheric pressure cold plasma on pathogenic oral biofilms and in vitro reconstituted oral epithelium. PLoS One. 2016;11:e0155427.
- [22] Kunhardt EE. Generation of large volume atmospheric pressure non equilibrium plasmas. IEEE Trans Plasma Sci. 2020;28:189-200.
- [23] Kogelschatz U. Filamentary, patterned, and diffuse barrier discharges. IEEE Trans Plasma Sci. 2002;30:1400-08.
- [24] Stoffels E, Flikweert AJ, Stoffels WW, Kroesen GM. Plasma needle: A nondestructive atmospheric plasma source for fine surface treatment of biomaterials. Plasma Sources Sci Technol. 2002;11:383-88.
- [25] Bernhardt T, Semmler ML, Schafer M, Bekeschus S, Emmert S, Boeckmann L. Plasma medicine: Applications of cold atmospheric pressure plasma in dermatology. Oxid Med Cell Longev. 2019;2019:3873928.
- [26] Cheng H, Xu J, Li X, Liu D, Lu X. On the dose of plasma medicine: Equivalent total oxidation potential (ETOP). Phys Plasmas. 2020;27:063514.
- [27] Pei X, Lu X, Liu J, Liu D, Yang Y, Ostrikov K, et al. Inactivation of a 25.5 µm Enterococcus faecalis biofilm by a room-temperature, battery-operated, handheld air plasma jet. J Phys D Appl Phys. 2012;45:165205.
- [28] Bonzanini AD, Shao K, Stancampiano A, Graves DB, Mesbah A. Perspectives on machine learning-assisted plasma medicine: Towards automated plasma treatment. IEEE Trans Radiation and Plasma Medical Sci. 2021. In Press.
- [29] Knipe DM, Howley PM, Gri_n DE, Lamb RA, Martin MA. Fields Virology, 5th ed.; Lippincott Williams & Wilkins: Philadelphia, PA, USA, 2006.
- [30] Weltmann KD, von Woedtke T. Plasma medicine- current state of research and medical application. Plasma Phys. Control. Fusion. 2017;59:014031.
- [31] Shen J, Tian Y, Li Y, Ma R, Zhang Q, Zhang J, et al. Bactericidal effects against s.aureus and physicochemical properties of plasma activated water stored at different temperatures. Sci Rep. 2016;6:28505.

- [32] Kamgang-Youbi G, Herry JM, Meylheuc T, Brisset JL, Bellon-Fontaine MN, Doubla A, et al. Microbial inactivation using plasma-activated water obtained by gliding electric discharges. Lett Appl Microbiol. 2009;48:13-18.
- [33] Lee K, Paek K, Ju WT, Lee Y. Sterilisation of bacteria, yeast, and bacterial endospores by atmospheric-pressure cold plasma using helium and oxygen. J Microbiol. 2006;44:269-75.
- [34] Mai-Prochnow A, Bradbury M, Ostrikov K, Murphy AB. Pseudomonas aeruginosa biofilm response and resistance to cold atmospheric pressure plasma is linked to the redox-active molecule phenazine. PLoS One. 2015;10:e0130373.
- [35] Tipa RS, Boekema B, Middelkoop E, Kroesen GMW. Cold plasma for bacterial inactivation. (2011).
- [36] Schairer DO, Chouake JS, Nosanchuk JD, Friedman AJ. The potential of nitric oxide releasing therapies as antimicrobial agents. Virulence. 2012;3:271-79.
- [37] Chauvin J, Judée F, Yousfi M, Vicendo P, Merbahi N. Analysis of reactive oxygen and nitrogen species generated in three liquid media by low temperature helium plasma jet. Sci Rep. 2017;7:4562.
- [38] Nicol MJ, Brubaker TR, Honish BJ, Simmons AN, Kazemi A, Geissel MA, et al. Antibacterial effects of low-temperature plasma generated by atmospheric-pressure plasma jet are mediated by reactive oxygen species. Sci Rep. 2020;10:3066.
- [39] Bourke P, Ziuzina D, Han L, Cullen PJ, Gilmore BF. Microbiological interactions with cold plasma. J Appl Microbiol. 2017;123:308-24.
- [40] Gilmore BF, Flynn PB, O'Brien S, Hickok N, Freeman T, Bourke P. Cold plasmas for biofilm control: Opportunities and challenges. Trends Biotechnol. 2018;36:627-38.
- [41] Zimmermann JL, Shimizu T, Schmidt HU, Li YF, Morfill GE, Isbary G. Test for bacterial resistance build-up against plasma treatment. New J Phys. 2012;14:073037.
- [42] Mai-Prochnow A, Clauson M, Hong J, Murphy AB. Gram positive and Gram negative bacteria differ in their sensitivity to cold plasma. Sci Rep. 2016;6:38610.
- [43] Vatansever F, de Melo WC, Avci P, Vecchio D, Sadasivam M, Gupta A, et al. Antimicrobial strategies centered around reactive oxygen species- bactericidal antibiotics, photodynamic therapy, and beyond. FEMS Microbiol Rev. 2013;37:955-89.
- [44] Kvam E, Davis B, Mondello F, Garner AL. Nonthermal atmospheric plasmarapidly disinfects multidrug-resistant microbes by inducing cell surface damage. Antimicrob Agents Chemother. 2012;56:2028-36.
- [45] Joshi SG, Paff M, Friedman G, Fridman G, Fridman A, Brooks AD. Control of methicillin-resistant Staphylococcus aureus in planktonic form and biofilms: A biocidal efficacy study of nonthermal dielectric-barrier discharge plasma. Am J Infect Contro. 2010;38(4):293-301.
- [46] Joshi SG, Cooper M, Yost A, Paff M, Ercan UK, Fridman G, et al. Nonthermal dielectric-barrier discharge plasma-induced inactivation involves oxidative DNA damage and membrane lipid peroxidation in Escherichia coli. Antimicrob Agents Chemother. 2011;55(3):1053-62.
- [47] Laroussi M, Mendis DA, Rosenberg M. Plasma interaction with microbes. New J Phys. 2003;5:41.
- [48] Costerton JW, Stewart PS, Greenberg EP. Bacterial biofilms: A common cause of persistent infections. Science. 1999;284:1318-22.
- [49] Høiby N, Bjarnsholt T, Givskov M, Molin S, Ciofu O. Antibiotic resistance of bacterial biofilms. Int J Antimicrob Agents. 2010;35:322-32.
- [50] Flemming HC, Wingender J. The biofilm matrix. Nat Rev Microbiol. 2010;8:623-33.
- [51] Abramzon N, Joaquin JC, Bray J, Brelles-Marino G. Biofilm destruction by RF high-pressure cold plasma jet. IEEE Trans Plasma Sci. 2006;34:1304-09.
- [52] Barraud N, Hassett DJ, Hwang SH, Rice SA, Kjelleberg S, Webb JS. Involvement of nitric oxide in biofilm dispersal of Pseudomonas aeruginosa. J Bacteriol. 2006;188:7344-53.
- [53] Holder D, Berry D, Dai D, Raskin L, Xi C. A dynamic and complex monochloramine stress response in Escherichia coli revealed by transcriptome analysis. Water Res. 2013;47:4978-85.
- [54] Kulkarni R, Antala S, Wang A, Amaral FE, Rampersaud R, Larussa SJ, et al. Cigarette smoke increases Staphylococcus aureus biofilm formation via oxidative stress. Infect Immun. 2012;80:3804-11.
- [55] Norton MD, Spilkia AJ, Godoy VG. Antibiotic resistance acquired through a DNA damage-inducible response in Acinetobacter baumannii. J Bacteriol. 2013;195:1335-45.
- [56] Ryu YH, Kim YH, Lee JY, Shim GB, Uhm HS, Park G, et al. Effects of background fluid on the efficiency of inactivating yeast with non thermal atmospheric pressure plasma. PLoS ONE. 2013;8:e66231.
- [57] Oehmigen K, Hoder T, Wilke C, Brandenburg R, Hahnel M, Weltmann KD, et al. Volume effects of atmospheric-pressure plasma in liquids. IEEE Trans Plasma Sci. 2011;39:2646-47.
- [58] Ikawa S, Kitano K, Hamaguchi S. Effects of pH on bacterial inactivation in aqueous solutions due to low-temperature atmospheric pressure plasma application. Plasma Process Polym. 2010;7:33-42.
- [59] Liu F, Sun P, Bai N, Tian Y, Zhou H, Wei S, et al. Inactivation of bacteria in an aqueous environment by a direct-current, cold-atmospheric-pressure air plasma microjet. Plasma Process Polym. 2010;7:231-36.
- [60] Oehmigen K, Hähnel M, Brandenburg R, Wilke C, Weltmann KD, von Woedtke T. The role of acidification for antimicrobial activity of atmospheric pressure plasma in liquids. Plasma Process Polym. 2010;7:250-57.
- [61] Roossinck MJ, Baz ER. Symbiosis: viruses as intimate partners. Annu Rev Virol. 2017;4:123-39.
- [62] Mehle, N. et al. Water-mediated transmission of plant, animal, and human viruses. In Advances in Virus Research (Malmstrom, C.M., ed.), 2018. pp. 85-128, Academic Press.
- [63] Staggemeier R, Bortoluzzi M, da Silva Heck TM, da Luz RB, Fabres RB, Soliman MC, et al. Animal and human enteric viruses in water and sediment samples from dairy farms. Agric Water Manag. 2015;152:135-41.

6

- [64] Zhang T, Breitbart M, Lee WH, Run JQ, Wei CL, Soh SW, et al. RNA viral community in human feces: Prevalence of plant pathogenic viruses. PLoS Biol. 2006;4:e3.
- [65] McMinn BR, Ashbolt NJ, Korajkic A. Bacteriophages as indicators of faecal pollution and enteric virus removal. Lett Appl Microbiol. 2017;65:11-26.
- [66] Cromeans T, Park GW, Costantini V, Lee D, Wang Q, Farkas T, et al. Comprehensive comparison of cultivable norovirus surrogates in response to different inactivation and disinfection treatments. Appl Environ Microbiol. 2014;80:5743-51.
- [67] Takamatsu T, Uehara K, Sasaki Y, Hidekazu M, Matsumura Y, Iwasawa A, et al. Microbial inactivation in the liquid phase induced by multi gas plasma jet. PLoS One. 2015;10:e0132381.
- [68] Lacombe A, Niemira BA, Gurtler JB, Sites J, Boyd G, Kingsley DH, et al. Nonthermal inactivation of norovirus surrogates on blueberries using atmospheric cold plasma. Food Microbiol. 2017;63:01-05.
- [69] Min SC, Roh SH, Niemira BA, Sites JE, Boyd G, Lacombe A. Dielectric barrier discharge atmospheric cold plasma inhibits Escherichia coli O157:H7, Salmonella, Listeria monocytogenes, and Tulane virus in Romaine lettuce. Int J Food Microbiol. 2016;237:114-20.
- [70] Park SY, Do Ha S. Assessment of cold oxygen plasma technology for the inactivation of major foodborne viruses on stainless steel. J Food Eng. 2018;223:42-45.
- [71] Nayak G, Aboubakr HA, Goyal SM, Bruggeman PJ. Reactive species responsible for the inactivation of feline calicivirus by a two-dimensional array of integrated coaxial microhollow dielectric barrier discharges in air. Plasma Process Polym. 2018;15:01-12.
- [72] Aboubakr HA, Mor SK, Higgins LA, Armien A, Youssef MM, Bruggeman PJ, et al. Cold argon-oxygen plasma species oxidize and disintegrate capsid protein of feline calicivirus. PLoS One. 2018;13:e0194618.
- [73] Aboubakr HA, Sampedro F, Collins JE, Bruggeman P, Goyal S. In situ inactivation of human norovirus GII.4 by cold plasma: Ethidium monoazide (EMA)- coupled RTqPCR underestimates virus reduction and fecal material suppresses inactivation. Food Microbiol. 2020;85:103307.
- [74] Bae SC, Park SY, Choe W, Ha SD. Inactivation of murine norovirus-1 and hepatitis A virus on fresh meats by atmospheric pressure plasma jets. Food Res Int. 2015;76:342-47.
- [75] Murray BK, Ohmine S, Tomer DP, Jensen KJ, Johnson FB, Kirsi JJ, et al. Virion disruption by ozone-mediated reactive oxygen species. J Virol Methods 2008;153(1):74-77.
- [76] Sakudo A, Toyokawa Y, Imanishi Y, Murakami T. Crucial roles of reactive chemical species in modification of respiratory syncytial virus by nitrogen gas plasma. Mater Sci Eng. 2017;74:131-36.
- [77] Toivonen L, Karppinen S, Schuez-Havupalo L, Teros-Jaakkola T, Mertsola J, Waris M, et al. Respiratory syncytial virus infections in children 0-24 months of age in the community. J Infect. 2019;80:69-75.
- [78] Mozetič M et al. Introduction to plasma and plasma diagnostics. In Non Thermal Plasma Technology for Polymeric Materials: Applications In Composites, Nanostructured Materials and Biomedical Fields (Thomas, S. et al., eds), 2019. pp. 23-65, Elsevier.
- [79] Wu Y, Liang Y, Wei K, Li W, Yao M, Zhang J, et al. MS2 virus inactivation by atmospheric pressure cold plasma using different gas carriers and power levels. Appl Environ Microbiol. 2015;81:996-1002.
- [80] Xia T, Yang M, Marabella I, Lee EM, Olson B, Zarling D, et al. Inactivation of airborne porcine reproductive and respiratory syndrome virus (PRRSv) by a packed bed dielectric barrier discharge non thermal plasma. J Hazard Mater. 2020;393:122266.
- [81] Wang G, Zhu R, Yang L, Wang K, Zhang Q, Su X, et al. Non thermal plasma for inactivated vaccine preparation. Vaccine. 2016;34:1126-32.
- [82] Su X, Tian Y, Zhou H, Li Y, Zhang Z, Jiang B, et al. Inactivation efficacy of nonthermal plasma activated solutions against Newcastle disease virus. Appl Environ Microbiol. 2018;84:e02836-17.
- [83] Kleinheksel A, Lee EM, Qiao Z, Wigginton KR, Clack HL. Inactivation of airborne viruses using a packed bed non thermal plasma reactor. J Phys D Appl Phys. 2019;52:255201.
- [84] Xia T, Kleinheksel A, Lee EM, Qiao Z, Wigginton KR, Clack HL. Inactivation of airborne viruses using a packed bed non thermal plasma reactor. J Phys D Appl Phys. 2019;52:255201.
- [85] Nicaise V. Crop immunity against viruses: Outcomes and future challenges. Front Plant Sci. 2014;5:660.
- [86] Lefeuvre P, Martin DP, Elena SF, Shepherd DN, Roumagnac P, Varsani A. Evolution and ecology of plant viruses. Nat Rev Microbiol. 2019;17:632-44.
- [87] Filipić A, Primc G, Zaplotnik R, Mehle N, Gutierrez-Aguirre I, Ravnikar M, et al. Cold atmospheric plasma as a novel method for inactivation of potato virus Y in water samples. Food Environ Virol. 2019;11:220-28.
- [88] Palou L, Valencia-Chamorro SA, Pérez-Gago MB. Antifungal edible coatings for fresh citrus fruit: A review. Coatings. 2015;5(4):962-86.
- [89] Hayashi N, Yagyu Y, Yonesu A, Shiratani M. Sterilisation characteristics of the surfaces of agricultural products using active oxygen species generated by atmospheric plasma and UV light. Japanese Journal of Applied Physics. 2014;53(5S1):05FR03.
- [90] Suhem K, Matan N, Nisoa M, Matan N. Inhibition of Aspergillus flavus on agar media and brown rice cereal bars using cold atmospheric plasma treatment. International Journal of Food Microbiology. 2013;161(2):107-11.
- [91] Avramidis G, Stüwe B, Wascher R, Bellmann M, Wieneke S, von Tiedemann A, et al. Fungicidal effects of an atmospheric pressure gas discharge and degradation mechanisms. Surface and Coatings Technology. 2010;205:S405-08.

- [92] Lee GJ, Sim GB, Choi EH, Kwon YW, Kim JY, Jang S, et al. Optical and structural properties of plasma-treated Cordyceps bassiana spores as studied by circular dichroism, absorption, and fluorescence spectroscopy. Journal of Applied Physics. 2015;117(2):023303.
- [93] Dasan BG, Mutlu M, Boyaci IH. Decontamination of Aspergillus flavus and Aspergillus parasiticus spores on hazelnuts via atmospheric pressure fluidized bed plasma reactor. International Journal of Food Microbiology. 2016;216:50-59.
- [94] Hashizume H, Ohta T, Fengdong J, Takeda K, Ishikawa K, Hori M, et al. Inactivation effects of neutral reactive-oxygen species on Penicillium digitatum spores using non equilibrium atmospheric-pressure oxygen radical source. Applied Physics Letters. 2013;103(15):153708.
- [95] Hashizume H, Ohta T, Takeda K, Ishikawa K, Hori M, Ito M. Oxidation mechanism of Penicillium digitatum spores through neutral oxygen radicals. Japanese Journal of Applied Physics. 2014;53(1):010209.
- [96] Hashizume H, Ohta T, Takeda K, Ishikawa K, Hori M, Ito M. Quantitative clarification of inactivation mechanism of Penicillium digitatum spores treated with neutral oxygen radicals. Japanese Journal of Applied Physics. 2015;54(1S):01AG05.
- [97] Iseki S, Hashizume H, Jia F, Takeda K, Ishikawa K, Ohta T, et al. Inactivation of Penicillium digitatum spores by a high-density ground-state atomic oxygenradical source employing an atmospheric-pressure plasma. Applied Physics Express. 2011;4(11):116201.
- [98] Eisenman HC, Casadevall A. Synthesis and assembly of fungal melanin. Applied Microbiology and Biotechnology. 2012;93(3):931-40.
- [99] Prokopowich D, Blank G. Microbiological evaluation of vegetable sprouts and seeds. J Food Prot. 1991;54:560-62.
- [100] Noriega E, Shama G, Laca A, Díaz M, Kong MG. Cold atmospheric gas plasma disinfection of chicken meat and chicken skin contaminated with Listeria innocua. Food Microbiol. 2011;28:1293-300.
- [101] Bußler S, Herppich WB, Neugart S, Schreiner M, Ehlbeck J, Rohn S, et al. Impact of cold atmospheric pressure plasma on physiology and flavonol glycoside profile of peas (Pisum sativum 'Salamanca'). Food Res Int. 2015;76:132-41.
- [102] Lacombe A, Niemira BA, Gurtler JB, Fan X, Sites J, Boyd G, et al. Atmospheric cold plasma inactivation of aerobic microorganisms on blueberries and effects on quality attributes. Food Microbiol. 2015;46:479-84.
- [103] Lee KH, Kim HJ, Yun SW, Jo C, Kim JK, Kim SH, et al. Evaluation of cold plasma treatments for improved microbial and physicochemical qualities of brown rice. LWT. 2016;73:442-47.
- [104] Pasquali F, Stratakos AC, Koidis A, Berardinelli A, Cevoli C, Ragni L, et al. Atmospheric cold plasma process for vegetable leaf decontamination: A feasibility study on radicchio (red chicory, Cichorium intybus L.). Food Control. 2016;60:552-59.
- [105] Sarangapani C, O'Toole G, Cullen P, Bourke P. Atmospheric cold plasma dissipation efficiency of agrochemicals on blueberries. Innov Food Sci Emerg Technol. 2017;44:235-41.
- [106] Ziuzina D, Misra NN, Cullen PJ, Keener K, Mosnier JP, Vilaró I, et al. Demonstrating the potential of industrial scale in-package atmospheric cold plasma for decontamination of cherry tomatoes. Plasma Med. 2016;6:397-412.
- [107] Whitehead JC. The Chemistry of Cold Plasma. In Cold Plasma in Food and Agriculture; Elsevier: Amsterdam, The Netherlands, 2016; pp. 53-81.
- [108] Misra N. Quality of Cold Plasma Treated Plant Foods. In Cold Plasma in Food and Agriculture; Elsevier: Amsterdam, The Netherlands, 2016; pp. 253-271.
- [109] Misra N, Jo C. Applications of cold plasma technology for microbiological safety in meat industry. Trends Food Sci Technol. 2017;64:74-86. [CrossRef].
- [110] Misra N, Schlüter O, Cullen PJ. Plasma in Food and Agriculture. In Cold Plasma in Food and Agriculture; Elsevier: Amsterdam, The Netherlands, 2016; pp. 1-16.
- [111] Niemira BA. Cold plasma decontamination of foods. Annu Rev Food Sci Technol. 2012;3:125-42.
- [112] Hung YW, Lee LT, Peng YC, Chang CT, Wong YK, Tung KC. Effect of a nonthermal-atmospheric pressure plasma jet on wound healing: An animal study. Journal of the Chinese Medical Association. 2016;79:320-28.
- [113] Fathollah S, Mirpour S, Mansouri P, Dehpour AR, Ghoranneviss M, Rahimi N, et al. Investigation on the effects of the atmospheric pressure plasma on wound healing in diabetic rats. Scientific Reports. 2016;6:19144.
- [114] Assadian O, Ousey KJ, Daeschlein G, Kramer A, Parker C, Tanner J, et al. Effects and safety of atmospheric low-temperature plasma on bacterial reduction in chronic wounds and wound size reduction: A systematic review and metaanalysis. International Wound Journal. 2019;16:103-11.
- [115] Brehmer F, Haenssle HA, Daeschlein G, Ahmed R, Pfeiffer S, Görlitz A, et al. Alleviation of chronic venous leg ulcers with a hand-held dielectric barrier discharge plasma generator (PlasmaDerm® VU-2010): Results of a monocentric, two-armed, open, prospective, randomized and controlled trial (NCT 01415622). Journal of the European Academy of Dermatology and Venereology. 2015;29:148-55.
- [116] Chuangsuwanich A, Assadamongkol T, Boonyawan D. The healing effect of low-temperature atmospheric-pressure plasma in pressure ulcer: A randomized controlled trial. The International Journal of Lower Extremity Wounds. 2016;15:313-19.
- [117] Lou BS, Lai CH, Chu TP, Hsieh JH, Chen CM, Su YM, et al. Parameters affecting the antimicrobial properties of cold atmospheric plasma jet. J Clin Med. 2019;8:930.
- [118] Kalghatgi S, Friedman G, Fridman A, Clyne AM. Endothelial cell proliferation is enhanced by low dose non thermal plasma through fibroblast growth factor-2 release. Annals of Biomedical Engineering. 2010;38:748-57.
- [119] Kisch T, Helmke A, Schleusser S, Song J, Liodaki E, Stang FH, et al. Improvement of cutaneous microcirculation by cold atmospheric plasma (CAP): Results of a controlled, prospective cohort study. Microvascular Research. 2016;104:55-62.
- [120] Giorgio M. Oxidative stress and the unfulfilled promises of antioxidant agents. Ecancermedicalscience. 2015;9:556.

- [121] Martinez L, Dhruv A, Lin L, Balaras E, Keidar, M. Interaction between a helium atmospheric plasma jet and targets and dynamics of the interface. Plasma Sources Science and Technology. 2019;28:115002.
- [122] von Woedtke T, Reuter S, Masur K, Weltmann KD. Plasmas for medicine. Phys Rep Rev Sect Phys Lett. 2013;530:291-320.
- [123] Dunnill C, Patton T, Brennan J, Barrett J, Dryden M, Cooke J, et al. Reactive oxygen species (ROS) and wound healing: The functional role of ROS and emerging ROS-modulating technologies for augmentation of the healing process. Int Wound J. 2017;14:89-96.
- [124] Ngo MHT, Liao JD, Shao PL, Weng CC, Chang CY. Increased fibroblast cell proliferation and migration using atmospheric N-2/Ar micro-plasma for the stimulated release of fibroblast growth factor-7. Plasma Proc Poly. 2014;11:80-88.
- [125] Ehlbeck J, Brandenburg R, von Woedtke T, Krohmann U, Stieber M, Weltmann KD. PLASMOSE antimicrobial effects of modular atmospheric plasma sources. GMS Krankenhhyg Interdiszip. 2008;3:Doc14.
- [126] Daeschlein G, Napp M, Lutze S, Arnold A, von Podewils S, Guembel D, et al. Skin and wound decontamination of multidrug-resistant bacteria by cold atmospheric plasma coagulation. J Dtsch Dermatol Ges. 2015;13:143-50. Doi: 10.1111/ddg.12559.
- [127] Moreau S, Moisan M, Tabrizian M, Barbeau J, Pelletier J, Ricard A, et al. Using the flowing afterglow of a plasma to inactivate Bacillus subtilis spores: Influence of the operating conditions, Journal of Applied Physics. 2000;88:1166.
- [128] Boudam MK, Moisan M, Saoudi B, Popovici C, Gherardi N, Massines F. Bacterial spore inactivation by atmospheric-pressure plasmas in the presence or absence of UV photons as obtained with the same gas mixture. Journal of Physics D: Applied Physics. 2006;39(16):3494-507.
- [129] Shimizu T, Steffes B, Pompl R, Jamitzky F, Bunk W, Ramrath K, et al. Characterisation of microwave plasma torch for decontamination. Plasma Processes and Polymers. 2008;5(6):577-82.
- [130] Lu XP, Ye T, Cao YG, Sun ZY, Xiong Q, Tang ZY, et al. The roles of the various plasma agents in the inactivation of bacteria. Journal of Applied Physics. 2008;104(5):53309.
- [131] Shi JJ, Kong MG. Cathode fall characteristics in a dc atmospheric pressure glow discharge. Journal of Applied Physics. 2003;94(9):5504-13.
- [132] Sladek REJ, Stoffels E, Walraven R, Tiebeek PJA, Koolhoven RA. Plasma treatment of dental cavities. IEEE Trans Plasma Sci. 2004;32:1540-43.
- [133] Goree J, Liu B, Drake D, Stoffels E. Killing of S. mutans bacteria using a plasma needle at atmospheric pressure. IEEE Trans Plasma Sci. 2006;34:1317-24.

- [134] Vandana BL. From distant stars to dental chairs: An update on plasma needle. Int J Dent Sci Res. 2014;2:19-20.
- [135] Yang B, Chen J, Yu Q, Li H, Lin M, Mustapha A, et al. Oral bacterial deactivation using a low-temperature atmospheric argon plasma brush. J Dent. 2011;9:48-56.
- [136] Dong X, Chen M, Wang Y, Yua Q. A mechanistic study of plasma treatment effects on demineralized dentin surfaces for improved adhesive/dentin interface bonding. Clin Plasma Med. 2014;2:11-16.
- [137] Ritts AC, Li H, Yu Q, Xu C, Yao X, Hong L, et al. Dentin surface treatment using a non thermal argon plasma brush for interfacial bonding improvement in composite restoration. Eur J Oral Sci. 2010;118:510.
- [138] Yavirach P, Chaijareenont P, Boonyawan D, Pattamapun K, Tunma S, Takahashi H, et al. Effects of plasma treatment on the shear bond strength between fiber reinforced composite posts and resin composite for core build-up. Dent Mater J. 2009;28:686-92.
- [139] Delben JA, Zago CE, Tyhovych N, Duarte S, Vergani CE. Effect of atmosphericpressure cold plasma on pathogenic oral biofilms and in vitro reconstituted oral epithelium. PLoS One. 2016;25:e0155427.
- [140] Koban I, Holtfreter B, Hübner NO, Matthes R, Sietmann R, Kindel E, et al. Antimicrobial efficacy of non thermal plasma in comparison to chlorhexidine against dental biofilms on titanium discs in vitro- Proof of principle experiment. J Clin Periodontol. 2011;38:956-65.
- [141] Jiang C, Chen MT, Gorur A, Schaudinn C, Jaramillo DE, Costerton JW, et al. Nanosecond pulsed plasma dental probe. Plasma Processes Polym. 2009;6:479-83.
- [142] Lee HW, Kim GJ, Kim JM, Park JK, Lee JK, Kim GC. Tooth bleaching with non thermal atmospheric pressure plasma. J Endod. 2009;35:587-91.
- [143] Nam SH, Lee HW, Cho SH, Lee JK, Jeon YC, Kim GC. High-efficiency tooth bleaching using non thermal atmospheric pressure plasma with low concentration of hydrogen peroxide. Journal of Applied Oral Science. 2013;21:265-70.
- [144] Lee HW, Nam SH, Mohamed AAH, Kim GC, Lee JK. Atmospheric pressure plasma jet composed of three electrodes: Application to tooth bleaching. Plasma Process Polym. 2010;7:274-80.
- [145] Park JK, Nam SH, Kwon HC, Mohamed AA, Lee JK, Kim GC. Feasibility of non thermal atmospheric pressure plasma for intracoronal bleaching. Int Endod J. 2011;44:170-75.
- [146] Kim MS, Koo IG, Choi MY, Jung JC, Eldali F, Lee JK, et al. Correlated electrical and optical studies of hybrid argon gas-water plasmas and their application to tooth whitening. Plasma Process Polym. 2012;8:339-45.

PARTICULARS OF CONTRIBUTORS:

- 1. Assistant Professor, Department of Mechanical Engineering, Alliance University, Bengaluru, Karnataka, India.
- 2. Assistant Professor, Department of Mechanical Engineering, Alliance University, Bengaluru, Karnataka, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR: Dr. G Divya Deepak, Chikkahadage Cross, Chandapura-Anekal, Main Road, Bengaluru-562106, Karnataka, India. E-mail: divyadeepak77@gmail.com

AUTHOR DECLARATION:

- Financial or Other Competing Interests: None
- Was informed consent obtained from the subjects involved in the study? No
- For any images presented appropriate consent has been obtained from the subjects. No

PLAGIARISM CHECKING METHODS: [Jain H et al.]

- Plagiarism X-checker: Jan 05, 2022
- Manual Googling: Feb 14, 2022
- iThenticate Software: Feb 22, 2022 (2%)

Date of Submission: Jan 03, 2022 Date of Peer Review: Jan 27, 2022 Date of Acceptance: Feb 18, 2022 Date of Publishing: Mar 01, 2022

ETYMOLOGY: Author Origin